

COARSE-GRAIN MODELLING OF LIPID BILAYERS: A LITERATURE REVIEW

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Received: 31st January 2007 / Published: 5th November 2007:

ABSTRACT

Particle-based computer simulation is a powerful tool to study the behaviour of membranes at molecular resolution. Atomic-level models have been employed for decades now, and have given an understanding of many membrane phenomena. However, these studies are computationally very expensive, for an enormous amount of calculation has to be performed to model the interactions between all atoms in the system. This problem can be tackled by adopting simplified, “coarse-grain” descriptions, in which the number of interacting particles is significantly reduced. In this review, we summarize and discuss the most representative work reported in the literature concerning coarse-grain computer models of lipid bilayers. Every model is analysed in terms of the force-field employed, parameterization procedure, and predictive power in relation to the corresponding experimental observables. We also highlight general advantages and drawbacks of the coarse-grain approach with respect to the traditional atomic-level methodology.

INTRODUCTION

Lipid bilayers are extremely complex systems: they are characterized by a highly heterogeneous structure and dynamics, and display an astonishingly rich and biologically-relevant behaviour on a wide range of spatial and temporal scales [1]. The scope of this article is limited to the nanosecond to microsecond temporal scale, and to the nanometre spatial scale, as this is the realm of the models that we will consider. The nature of the modelling methodology further limits the treatment to *non-specific* membrane phenomena, in the sense that biochemical reactions will not be involved. It is important to note that these restrictions do not compromise in any way the possibility of obtaining a general understanding of bilayers: experiments have indeed shown that the most fundamental membrane properties depend exclusively on basic *physical* principles governing lipid structure and dynamics *at the molecular level* [2].

Experimental investigation of the physics of membranes has resulted in the characterization of a large body of bilayer features. The internal structure is resolved via electron density profiles [3], lipid dynamics is studied by lateral diffusion measurements [4], mechanical properties can be related to the measurements of elastic moduli [5], electrostatic properties can be quantified by estimating internal potentials [6], and even the trans-bilayer stress distribution can be qualitatively measured [7]. All of these membrane properties are crucially important in a vast number of physiological mechanisms. For example, the thickness of the inner hydrophobic core directly influences the conformation and function of embedded proteins [8], whereas lipid fluidity is central to membrane lateral organization [9] and trans-membrane transport processes [10]. The bilayer dipole potential is believed to affect the behaviour of integral proteins [11], membrane fusion [12,13] and the modulation of molecule-membrane interactions in lipid rafts with effects on cells signalling [14, 15]. An even more important role is assigned to the trans-membrane lateral stress distribution, which is involved in protein folding [16], lipid synthesis [17], phase transitions and fusion [18–20] bilayer permeability [21], drug transport [22], anaesthetic potency [23–25] and modulation of protein channels [26–29].

Considering the hugely differing nature of the structures, interactions and phenomena present in lipid bilayers, it is clear that the development of realistic membrane models is a challenging task. In this review, we focus on particle-based computer models, which are typically simulated with the molecular dynamics method [30].

In particular, we will describe the two main approaches to the simulation of membranes. First, we summarize the basics of the traditional methodology, which involves an *atomistically-detailed* description of the system. Second, we describe the recently developed *coarse-grain* approach, which is based on a simplified representation: this is the main focus of this review, and therefore a number of representative coarse-grain models that have appeared in the literature will be described in some detail.

ATOMIC-LEVEL MODELLING

The traditional methodology employed to simulate lipid bilayers is based on an atomic-level (AL) representation: every atom of the system is explicitly represented as a point-mass. The inter-atomic interactions are described by molecular mechanics force-fields [31]: non-bonded interactions are modelled using Lennard-Jones and Coulomb potentials, whereas bond terms are considered via harmonic, angle and dihedral potentials.

Over the past two decades, several AL models of lipid bilayers have been developed and validated on experimental structural and dynamic data [32–39]. More recently, AL membrane simulations have been used to study important biological phenomena, such as undulations [40], self-diffusion [41], electrostatic interactions [42, 43], cholesterol function [44, 45], the permeation of small molecules [46, 47], the lateral pressure distribution [48–50] and the appearance of transient ordered domains [51]. Despite their popularity, AL membrane models are affected by a limiting efficiency issue: obtaining data comparable to the experimental measurements is hugely time-consuming, due to the computational cost inherent in the simulation of every atom in the system. This drawback of the AL approach makes difficult the study of (relatively) large-scale phenomena, such as membrane fusion or lipid rafts. Furthermore, there are efficiency issues associated even with the standard (relatively small) membrane sizes. For instance, it has become clear that several tens of nanoseconds are required for some crucial parameters (such as the lipid area) to converge [52]: even with parallel computing, several weeks of computation may be needed just to equilibrate the AL system. Also, the measurement of some properties such as elastic moduli or the lateral pressures, typically require extended simulation times to obtain accurate and precise data. Another issue is related to the prohibitive cost of simulating the fundamental phenomenon of the self-assembly of lipids into a bilayer from a random solution. With the single exception of Marrink *et al.* [53], all AL studies reported in the literature to date have been conducted on pre-assembled bilayers: these systems are therefore not guaranteed to be thermodynamically stable. A more general problem concerns the force-field reliability: AL lipid force-fields are still far from accurate, in terms of being able to *quantitatively* reproduce experimental data. For example, Benz *et al.* [54] conducted a thorough testing on the popular CHARMM and GROMACS force-fields via constant-pressure simulation, and concluded that neither parameter set can capture within experimental error the experimentally determined structure of a DOPC bilayer in the fluid state. Furthermore, de Vries *et al.* [55] showed through constant-volume simulation of a standard hydrated DPPC bilayer that the GROMOS parameter set yields a total pressure of about -140 bar (the proper equilibrium value being 1 bar). A refinement of the current lipid force-fields could in principle be performed by trial-and-error adjustments of the parameters, although in practice the computational cost associated with this procedure may be too high.

COARSE-GRAIN MODELLING

A possible solution to the AL efficiency problem involves the use of simpler, *coarse-grain* (CG) models. The general CG strategy involves grouping together selected clusters of atoms into single super-sites, to reduce the number of interactions calculated, and hence

also the computational cost (as the computing time scales with the square of the number of interaction sites present). A typical lipid molecule, which in reality comprises more than one hundred atoms, is thus typically reduced to a collection of only a dozen CG sites. The representation of water also undergoes a simplification process, that typically results in the replacement of groups of three/four water molecules by single CG units. To further improve efficiency, electrostatic interactions are highly simplified or completely absent.

In the following sections, we review the most representative coarse-grain models for biological membranes that have been reported in the literature, in chronological order. We focus on those CG models that retain a connection with specific chemical systems: these studies allow a direct comparison with the experimental results for the corresponding bilayer systems. However, a section is also devoted to a brief summary of “coarser”, idealized models: in this case, there is no effort to reproduce any particular real system, the objective instead being to capture general membrane phenomena (see *Idealized membrane models*). Other reviews of CG biomembrane modelling have been published by Nielsen *et al.*[56], Shillcock and Lipowsky [57], Muller *et al.*[58] and Venturoli *et al.*[59].

SMIT AND GROOT MODELS

In the early Nineties, Smit and co-workers developed a seminal CG model of oil/water/surfactant systems [60, 61]: simulations showed for the first time the spontaneous formation of micelles. Some years later, Groot and Warren [62] discussed the use of the Dissipative Particle Dynamics (DPD) technique to simulate the dynamics of mesoscopic systems, and also proposed parameterization methods. The DPD technique is a coarse-grain scheme by construction: the forces due to clusters of individual molecules are lumped together to yield effective friction and a fluctuating force between the interacting sites. In particular, beads interact pairwise via a combination of three contributions: a conservative-repulsive, a dissipative, and a random force [63]. On these bases, Venturoli and Smit [64] employed DPD to model single-chain surfactant bilayers while Groot [65] simulated the spontaneous formation of surfactant micelles and the formation of polymer-surfactant mixtures. In all these early studies however, the parameters had not been related to molecules of specific chemistry. This issue was addressed by Groot and Rabone [66], who developed a model for phosphatidylethanolamine (PE) membranes; they also included similar models of non-ionic surfactants. The parameterization method of Groot and co-workers was subsequently used and extended by Smit and co-workers to develop a model of dimyristoylphosphatidylcholine (DMPC) lipids in water [67].

Force-field and parameterization

Smit *et al.* [61] modelled idealized surfactant molecules as chains of identical Lennard-Jones sites.

Venturoli and Smit [64] employed different parameters to construct surfactants as linear molecules composed of one headgroup (hydrophilic) site and six to ten tail (hydrophobic) sites. Groot and Rabone [66] parameterized a DPD force-field to represent PE lipids. In

particular, triplets of methyl groups were coarse-grained into individual tail beads, whereas the glycerol-headgroup region was described by three CG sites. Parameters were derived from compressibility and solubility data. Triplets of water molecules were collected into individual DPD solvent beads, the parameters being fitted to give the correct compressibility of water. Many subsequent DPD studies have employed this model of solvation [66].

A model of DMPC consisting of three hydrophilic head beads and two tails, each consisting of five hydrophobic tail beads was implemented by Kranenburg *et al.*[67]. The chain length of this model was varied by Kranenburg and Smit [68] to model the dilaureoylphosphatidylcholine (DLPC) and distearoylphosphatidylcholine (DSPC) lipid types. A common feature of the Smit and Groot CG membrane force-fields is the complete lack of electrostatic interactions. Groot [69] did extend the DPD methodology to incorporate electrostatics, obtaining realistic results for a cationic polyelectrolyte solution, but no biomembrane model has been developed to date.

Results

The very simple oil/water/surfactant model of Smit *et al.* [61] was able to (qualitatively) reproduce experimental observations such as micelle formation, density profiles and order parameters.

Venturoli and Smit [64] obtained for the first time the self-assembly of (model) surfactant bilayers via DPD simulations. Studies on the effect of changes in the chain length and stiffness of the single-tail surfactants on the properties of the model membranes were carried out. The lateral stress profile across the model bilayer was also computed: the distribution is qualitatively reasonable. Groot and Rabone [66] made the first attempt to simulate a realistic biomembrane with DPD using a molecular-specific parameterization: the resulting CG membrane structure matched that of corresponding AL simulations, and the essential experimental thermodynamics was reproduced. They also included non-ionic surfactants to investigate morphology changes and bilayer rupture processes.

Smit and coworkers worked on further tuning of Groot's parameter set. Kranenburg *et al.* [70] studied the phase behaviour of model membranes and were able to induce interdigitation; the self-assembly process was also simulated [71]. Kranenburg *et al.* [67, 72] reproduced the experimental values of area per lipid and the hydrophobic thickness of a DMPC bilayer. The phase behaviour of bilayers comprising different lipid species was also characterized [68].

Kranenburg and Smit [73] also developed a DPD system composed of a DSPC bilayer incorporating model alcohol molecules, represented as idealized amphiphiles. The study reproduced the experimental phase diagrams, as well as the alcohol-induced interdigitated phase shown by experiments [74]. Smit's model has also been extended to study the structural changes resulting from the inclusion of a rod-like object taken as an idealized protein [75].

KLEIN MODEL

Klein and coworkers developed a model for simulating hydrated DMPC lipid bilayers which for the first time included an explicit, though incomplete, treatment of electrostatic interactions [76].

Force-field and parameterization

In the Klein CG model, the 118 atoms of a DMPC lipid are reduced to a 13-site model. The two choline and phosphate head-groups were assigned charges of $+e$ and $-e$, respectively: these are the only charges present in the force-field, and they interact via a dielectric constant set to $\epsilon_r=78$. The potentials employed were systematically parameterized in order to mimic structural properties obtained from atomistic simulations (radial distribution functions) and experimental data (density, surface tension).

Klein *et al.* modelled water through spherically symmetric sites each representing a loose grouping of three water molecules; site-site interactions were described using a Lennard-Jones potential, the parameters being chosen to reproduce the correct density and to have consistency with hydrodynamics. The electrostatic properties of water were not included in the solvent model [76].

Results

Shelley *et al.* [77] qualitatively reproduced the density profiles of a hydrated liquid-phase DMPC bilayer; simulations were conducted at constant-volume. Lopez *et al.* [78] further simulated Klein's model and studied the lipid lateral diffusion: the CG diffusion coefficient was about two orders of magnitude higher than the experimental measurement.

The Klein model has been extended to incorporate the anaesthetic halothane inside the DMPC bilayer [77, 79]. A single site was used to represent each halothane molecule. The system studied was characterized by a 2:1 ratio of the phospholipid to halothane, equivalent to an atomistic simulation considered for comparison. After several adjustments of the parameters, the distribution of the halothane perpendicular to the membrane was brought into qualitative agreement with that found in the atomistic studies. Pickholz *et al.* [79] observed that increasing the anaesthetic concentration resulted in an increase of the lipid area and order parameters and a decrease in the inter-lamellar spacing.

Srinivas and Klein [80] studied the interaction of a synthetic pore-promoting "hydraphile" molecule with the CG phospholipid bilayer; the system was simulated for 5 ns, during which the initially fully extended trans-membrane hydraphile adjusted its end-to-end distance to match the bilayer thickness.

Srinivas *et al.* [81] employed Klein's CG model to simulate diblock copolymer self-assembly.

The Klein bilayer was also extended to incorporate a model nanotube [56, 82]. In particular, the nanotube is modelled as a hydrophobic rod (made of sites identical to the lipid tail sites) capped at its termini with hydrophilic sites (identical to the water sites). Simulations were carried out in the NPT ensemble for several tens of nanoseconds to study the structure and dynamics of spontaneous insertion into the CG membrane [83]. First the nanotube fuses with a bilayer leaflet, then penetrates the interior while rotating to assume a transverse orientation.

Nielsen *et al.* [84] studied the lipid bilayer perturbations around the trans-membrane nanotube, focusing on the contact angle at the bilayer-nanotube interface and on the orientation of the lipid molecules in the vicinity of the inclusion.

Nielsen *et al.* [85] studied the trans-membrane peptide-induced lipid sorting (the phenomenon by which integral proteins attract the lipid type which better matches their hydrophobic surface) and the mechanism of L-to-inverted phase transition.

The spontaneous insertion of antimicrobial polymers has also been simulated: the average orientation of the antimicrobial molecules was found to be parallel to the bilayer plan [86].

MARRINK MODEL

Marrink *et al.* [87] developed a CG model for lipid simulation which has become very popular due to its high efficiency, flexibility and simplicity.

Force-field and parameterization

In the Marrink model, DPPC molecules were coarse-grained into 12 sites [87]. Water is represented by Lennard-Jones sites accounting for groups of four water particles. A trial and error procedure was used to optimize the parameters to reproduce the experimental densities of pure water and alkane systems around room temperature, the mutual solubility of oil and water, and the relative diffusion rates. Electrostatic interactions are only present between lipid headgroups, where they are treated through a Coulombic potential in a manner similar to the Klein model. A relative dielectric constant $\epsilon_r = 20$ is employed for explicit screening [87].

Results

Marrink's model showed spontaneous bilayers formation: the final structures were consistent with corresponding AL results, in terms of density profiles and order parameters. A number of elastic parameters were computed: results for the area compressibility modulus, bending rigidity and line tension were within an order of magnitude of the experimental data [87]. The calculation of the bending rigidity modulus proved particularly demanding: the simulation of undulatory modes (from which the bending rigidity can be extracted)

required the simulation of a bilayer patch comprising 6400 lipids for 250 ns. The lipid lateral diffusion coefficient was also computed: it was found to be about four times larger than the experimental data [87].

The phase transformation into the gel phase was simulated by Marrink *et al.* [88]: the observed drop in lateral mobility by two orders of magnitude is consistent with experiments.

The Marrink CG model has been used to simulate a variety of phenomena and extended-systems, such as vesicle formation [89] and fusion [90], and mixed-lipid systems [91].

By increasing the temperature or reducing the hydration level, Marrink and Mark [92] simulated the complete transition pathway from a multi-lamellar to an inverted hexagonal phase: stalk intermediates were identified, in agreement with experimental observation.

Faller and Marrink [93] studied binary mixtures of two phosphatidylcholines of different chain lengths: the experimental phase behaviour was qualitatively reproduced. Shi and Voth [94] also employed Marrink's methodology to simulate a binary mixed system at the liquid-gel phase coexistence condition.

Dickey and Faller [95] studied the interaction of Marrink's bilayer with an alcohol. The Marrink model has also been extended to contain CG representations of polyamidoamine dendrimers [96] and membrane proteins [97, 98].

Recently, Adhangale and Gaver [99] employed the Marrink model to study a DPPC monolayer at the air/water interface.

VOTH MODEL

Izvekov and Voth [100] developed a CG model for hydrated DMPC bilayers using a multi-scale approach in which explicit atomistic forces are propagated in scale to the coarse-grained level.

Force-field and parameterization

The Voth model relies on a force-field which is obtained from a corresponding AL simulation via a so-called “force-matching” procedure. In particular, the force-matching yields potentials of mean force (PMFs) that are fitted using a spline interpolation of the AL forces, where atoms are grouped according to the chosen CG subdivision. This method is unique in the CG field, as it is not dependent on the matching of selected thermodynamic data, but it makes use of the calculated atomic forces from an underlying AL model.

Izvekov and Voth coarse-grained each DMPC lipid to a 13-site model, in a way similar to the Klein and Marrink models. Water molecules are represented explicitly through individual CG sites. Electrostatic interactions are not present in this CG force-field.

Results

A pre-assembled membrane bilayer comprising 64 DMPC molecules solvated by 1312 water sites was simulated in the NVT ensemble. Radial distribution functions and density profiles of the underlying AL model were reproduced [100].

The “force-matching” procedure has subsequently been applied to the study of a DMPC/cholesterol mixture simulated at constant temperature and pressure [101]. CG order parameters were consistent with the underlying AL data. CG diffusion coefficients turned out to be about four times higher than the corresponding experimental measurements.

Shi *et al.* [102] constructed a mixed AL-CG model of a membrane-protein system: in particular, an AL model of the gramicidin A ion channel was embedded in hydrated CG lipid bilayer. The system was simulated at constant temperature and volume for 10 ns; the radial distribution functions of the AL simulation employed in the “force-matching” parameterization were reproduced.

IDEALIZED MEMBRANE MODELS

This section briefly presents CG models that are more phenomenological and simplistic than those previously summarized: bilayers are composed of amphiphiles with no specific chemical identity. These models are sometimes called “bead-and-spring” models, as the interactions represented often reduce to Lennard-Jones and harmonic potentials only. Electrostatic interactions are indeed typically not included. In some cases, even the solvent is sacrificed (see *Implicit-solvent models*).

LIPOWSKY MODEL

In the late Nineties, Lipowsky and coworkers developed a simple, idealized CG bilayer model that proved capable of qualitatively capturing a number of fundamental membrane characteristics [103]. In particular, solvated aggregates of surfactant molecules were obtained employing only two types of Lennard-Jones sites: hydrophilic sites, used to describe both solvent and surfactant head-group particles, and hydrophobic sites, employed to model surfactant tail segments. Simulations of the model allowed bilayer self-aggregation, diffusion, interfacial tension and area compressibility to be studied. The trans-bilayer lateral pressure profile was also calculated: the distribution is qualitatively reasonable, apart from the unphysical negative pressure peak at the bilayer mid-plane. The Lipowsky model was further investigated in terms of undulations: in particular, from the spectral analysis of the bilayer shape fluctuations, Goetz *et al.* [104] were able to extract a value for the bending rigidity modulus.

Imparato *et al.* [105] extended the model to simulate a mixture of two different types of molecules: lateral diffusion was measured, and the activation barrier of flip-flop processes was estimated. Imparato *et al.* [106] investigated further the two-component membrane to study shape fluctuations and elastic properties. The Lipowsky model was employed by den

Otter and Briels [107] to study the bilayer thermally-induced undulations and to ultimately extract the bending rigidity using different methods: undulatory modes either aroused naturally during equilibrium simulations, or were imposed through a number of non-equilibrium methods. The results of this work have proved consistent with the bending free energy prediction from the popular theory developed by Helfrich [108]. Boek *et al.* [109] performed additional simulations on Lipowsky's model to analyse structure factors: they propose refining the intermolecular potential parameters to yield fluctuation spectra that coincide with the atomistic results.

OTHER BEAD-AND-SPRING MODELS

Loison *et al.* [110] proposed a model where amphiphiles are represented by linear tetramers, each composed of two solvophobic (tail) and two solvophilic (head) beads. A self-assembled bulk lamellar phase was investigated from a mechanical perspective. Pores, fluctuations and defects of the lamellar stack were also analysed [111, 112].

Stevens [113] developed a CG model where each lipid is composed of eleven beads, four in each aliphatic tail and three for the headgroup; the solvent is represented by particles equivalent to the headgroup sites. Stevens' model system spontaneously self-assembled into a lamellar bilayer; measurements of lipid lateral diffusion and material elastic parameters yielded qualitatively reasonable values [113]. Fusion simulations have also been performed: at the contact edge between liposomes, lipids were observed to splay their tails into the opposing leaflets, thus progressively producing a new hydrophobic core (a “stalk”) that eventually opened to complete the fusion process [114].

Lenz and Schmid [115] presented a simple CG bead-spring model with the aim of reproducing the main (gel-liquid) phase transition of biomembranes. Lipids are modelled as single-tail amphiphiles, each composed of six tail beads and one slightly larger head bead. Molecules are defined via bond and bond-angle harmonic potential; non-bonded interactions are taken into account via Lennard-Jones potentials. Solvent was represented by explicit “phantom” solvent beads: they behave exactly like head beads, except that they do not interact with each other. The model proved well suited for simulating lipid bilayers in the regime of the liquid-gel transition.

IMPLICIT-SOLVENT MODELS

In this section we briefly consider a number of CG membrane models that have been designed to work without the presence of explicit solvent; for a thorough review, see Brannigan *et al.*[116].

Although water is generally regarded as an essential component of bilayer systems [2], in specific cases it may not play a fundamental role while still representing a computational bottle-neck. Hence, solvent-free models might be useful to gain insight into selected membrane phenomena at a very low computational cost.

Whitehead *et al.* [117] developed a CG model for the bilayer hydrocarbon region only, based on the Gay-Berne model of liquid crystals [118]: in the absence of water and head-group, the lipid packing was maintained through the use of a restrain potential. The experimental phase behaviour of typical phospholipid bilayers was reproduced.

Noguchi and Takasu [119] represented lipids as rigid trimers consisting of one hydrophilic and two hydrophobic units interacting through multi-body potentials. This model has been used to simulate the self-assembly [119] and fusion [120] of vesicles, adhesion of nanoparticles to vesicles [121] and pulled vesicles [122].

Lyubartsev [123] developed a 10-site lipid model parameterized exclusively from corresponding AL structural data processed through the inverse Monte Carlo method [124]. Several simulations of the CG model obtained were performed, both within Monte Carlo and molecular dynamics simulations, such as a periodic sample of lipid molecules ordered in a bilayer, a free sheet of such a bilayer without periodic boundary conditions, formation of vesicle from a plain membrane and self-assembly of lipids [123].

Brannigan and Brown [125] developed a model of bilayers where entire lipids are represented by single soft spherocylinder. Through the combination of three simple pair potentials, a rich assortment of self-assembled phases was recovered, including micelles, fluid bilayers and gel-like bilayers. Brannigan *et al.* [126] extended the model to allow for variations in lipid length and simulations under constant surface tension conditions.

Brannigan *et al.* [127] developed a new model with flexible lipids that was employed to extract compressibility and bending moduli, and the lateral pressure profile. The pressure distribution is qualitatively reasonable apart from an unrealistic pronounced pressure trough located at the centre of the bilayer; interestingly, the curve is very similar to the result from the explicit-solvent model by Goetz and Lipowsky [103].

The model by Brannigan *et al.* [127]) was extended to incorporate multiple lipid species: in particular, the elasticity of idealized heterogeneous bilayers has been analysed [128].

SUMMARY AND CONCLUSION

The traditional methodology for simulating biomembranes involves an atomic-level description of lipids and hydrating water. This approach has yielded fairly precise and accurate predictions of a number of experimental data. However, the associated large computational cost results in two major issues:

- collective, large-scale phenomena such as membrane fusion or micro-domain formation (lipid rafts) are intractable;
 - the reliability of the calculation of some important properties, such as the lipid area or the lateral pressure profile, is often undermined by insufficient sampling.
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Coarse-grain modelling techniques have shown a number of advantages over atomistic models:

- orders of magnitude more efficient, resulting in the possibility to study phenomena characterized by much larger temporal and spatial scales;
- enhanced sampling, resulting in statistically-reliable measurements;
- bilayers are usually self-assembled, which guarantees that the system is at thermodynamic equilibrium.

On the other hand, CG models are also affected by issues:

- limited force-field transferability;
- experimental data are typically captured at a qualitative level only;
- the over-simplification or complete lack of electrostatics precludes the proper representation of the membrane potential, which has indeed never been computed for CG models;
- no CG model published to date has proved able to reasonably quantify the trans-bilayer lateral pressure profile, which is crucial for the accurate modelling of many membrane processes.

There is a clear need, therefore, for CG models to be developed that incorporate electrostatic interactions more accurately. Such models will likely increase the range of membrane phenomena that may be accurately studied using the CG approach.

Considering that the CG field is still in its infancy, the preliminary results obtained so far are encouraging. In the future, we foresee that coarse-grain models will play an increasingly important role in the understanding of lipid membranes.

REFERENCES

- [1] Lipowsky, R., Sackmann, E. (1995) *Structure and Dynamics of Membranes*. Elsevier.
 - [2] Mouritsen, O.G. (2005) *Life – As a Matter of Fat. The Emerging Science of Lipidomics*. First edn. Springer.
 - [3] Nagle, J.F., Tristram-Nagle, S. (2000) Structure of lipid bilayers. *Biochim. Biophys. Acta* **1469**:159 – 195.
 - [4] Filippov, A., Oradd, G., Lindblom, G. (2003) The effect of cholesterol on the lateral diffusion of phospholipids in oriented bilayers. *Biophys. J.* **84**:3079 – 3086.
-

- [5] Rawicz, W., Olbrich, K.C., McIntosh, T., Needham, D., Evans, E. (2000) Effect of chain length and unsaturation on elasticity of lipid bilayers. *Biophys. J.* **79**:328 – 339.
 - [6] Wang, L., Bose, P.S., Sigworth, F.J. (2006) Using cryo-EM to measure the dipole potential of a lipid membrane. *Proc. Natl Acad. Sci. U S A* **103**:18528.
 - [7] Templer, R.H., Castle, S.J., Curran, A.R., Rumbles, G., Klug, D.R. (1998) Sensing isothermal changes in the lateral pressure in model membranes using di-pyrenyl phosphatidylcholine. *Faraday Discuss.* **111**:41 – 53.
 - [8] Bezrukov, S.M. (2000) Functional consequences of lipid packing stress. *Curr. Opin. Colloid In.* **5**:237 – 243.
 - [9] Bagatolli, L.A. (2006) To see or not to see: lateral organization of biological membranes and fluorescence microscopy. *Biochim. Biophys. Acta* **1758**:1541 – 1556.
 - [10] Bloom, M., Evans, E., Mouritsen, O.G. (1991) Physical properties of the fluid lipid-bilayer component of cell membranes: a perspective. *Q. Rev. Biophys.* **24**:293 – 397.
 - [11] Clarke, R.J. (2001) The dipole potential of phospholipid membranes and methods for its detection. *Adv. Colloid Interface Sci.* **89**:263 – 281.
 - [12] Cladera, J., Martin, I., Ruyschaert, J.M., O'Shea, P. (1999) Characterization of the sequence of interactions of the fusion domain of the simian immunodeficiency virus with membranes-Role of the membrane dipole potential. *J. Biol. Chem.* **274**:29951 – 29959.
 - [13] Cladera, J., Martin, I., O'Shea, P. (2001) The fusion domain of HIV gp41 interacts specifically with heparan sulfate on the T-lymphocyte cell surface. *Embo J.* **20**:19-26.
 - [14] Luker, G.D., Flagg, T.P., Sha, Q., Luker, K.E., Pica, C.M., Nichols, C.G., Piwnicka-worms, D. (2001) MDR1 P-glycoprotein reduces influx of substrates without affecting membrane potential. *J. Biol. Chem* **276**:49053-49060.
 - [15] O'Shea, P. (2003) Intermolecular interactions with/within cell membranes and the trinity of membrane potentials: kinetics and imaging. *Biochem. Soc. T.* **31**:990 – 996.
 - [16] Curran, A.R., Templer, R.H., Booth, P.J. (1999) Modulation of folding and assembly of the membrane protein bacteriorhodopsin by intermolecular forces within the lipid bilayer. *Biochemistry* **38**:9328 – 9336.
 - [17] Attard, G.S., Templer, R.H., Smith, W.S., Hunt, A.N., Jackowski, S. (2000) Modulation of CTP: phosphocholine cytidyltransferase by membrane curvature elastic stress. *Proc. Natl Acad. Sci. U S A* **97**:9032 – 9036.
 - [18] Gruner, S.M. (1985) Intrinsic curvature hypothesis for biomembrane lipid composition: a role for nonbilayer lipids. *Proc. Natl Acad. Sci. U S A* **82**:3665 – 3669.
-

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- [19] Siegel, D.P., Kozlov, M.M. (2004) The Gaussian curvature elastic modulus of N-nonomethylated dioleoylphosphatidylethanolamine: Relevance to membrane fusion and lipid phase behavior. *Biophys. J.* **87**:366–374.
- [20] Kozlovsky, Y., Efrat, A., Siegel, D.P., Kozlov, M.M. (2004) Stalk phase formation: effects of dehydration and saddle splay modulus. *Biophys. J.* **87**:3619–3619.
- [21] Xiang, T.X., Anderson, B.D. (1994) Molecular distributions in interphases: statistical mechanical theory combined with molecular dynamics simulation of a model lipid bilayer. *Biophys. J.* **66**:561–572.
- [22] Curnow, P., Lorch, M., Charalambous, K., Booth, P.J. (2004) The reconstitution and activity of the small multidrug transporter EmrE is modulated by non-bilayer lipid composition. *J. Mol. Biol.* **343**:213–222.
- [23] Cantor, R.S. (1997) The lateral pressure profile in membranes: A physical mechanism of general anesthesia. *Biochemistry* **36**:2339–2344.
- [24] Cantor, R.S. (2001) Breaking the Meyer-Overton rule: Predicted effects of varying stiffness and interfacial activity on the intrinsic potency of anesthetics. *Biophys. J.* **80**:2284–2297.
- [25] Mohr, J.T., Gribble, G.W., Lin, S.S., Eckenho, R.G., Cantor, R.S. (2005) Anesthetic potency of two novel synthetic polyhydric alkanols longer than the n-alkanol cutoff: Evidence for a bilayer-mediated mechanism of anesthesia? *J. Med. Chem.* **48**:4172–4176.
- [26] Keller, S.L., Bezrukov, S.M., Gruner, S.M., Tate, M.W., Vodyanoy, I., Parsegian, V.A. (1993) Probability of alamethicin conductance states varies with nonlamellar tendency of bilayer phospholipids. *Biophys. J.* **65**:23–27.
- [27] Perozo, E., Kloda, A., Cortes, D.M., Martinac, B. (2002) Physical principles underlying the transduction of bilayer deformation forces during mechanosensitive channel gating. *Nature Struct. Biol.* **9**:696–703.
- [28] Jensen, M.O., Mouritsen, O.G., Peters, G.H. (2004) Simulations of a membrane-anchored peptide: structure, dynamics, and influence on bilayer properties. *Biophys. J.* **86**:3556–3575.
- [29] van den Brink-van der Laan, E., Killian, J.A., de Kruij, B. (2004) Nonbilayer lipids affect peripheral and integral membrane proteins via changes in the lateral pressure profile. *Biochim. Biophys. Acta* **1666**:275–288.
- [30] Rapaport, D.C. (2004) *The Art of Molecular Dynamics Simulation*, 2nd edn. Cambridge University Press, Cambridge.
- [31] Leach, A.R. (2001) *Molecular Modelling – Principles and Applications*, 2nd edn. Prentice Hall.
-

- [32] van der Ploeg, P., Berendsen, H.J.C. (1982) Molecular dynamics simulation of a bilayer membrane. *J. Chem. Phys.* **76**:3271 – 3276.
- [33] van der Ploeg, P., Berendsen, H.J.C. (1983) Molecular dynamics of a bilayer membrane. *Mol. Phys.* **49**:233-248.
- [34] Egberts, E., Marrink, S.-J., Berendsen, H.J.C. (1994) Molecular dynamics simulation of a phospholipid membrane. *Eur. Biophys. J.* **22**:423 – 436.
- [35] Essex, J.W., Hann, M.M., Richards, W.G. (1994) Molecular dynamics simulation of a hydrated phospholipid bilayer. *Phil. Trans. R. Soc. Lond. B.* **344**:239 – 260.
- [36] Tieleman, D.P., Marrink, S.-J., Berendsen, H.J.C. (1997) A computer perspective of membranes: molecular dynamics studies of lipid bilayer systems. *Biochim. Biophys. Acta* **1331**:235 – 270.
- [37] Merz, K.M. (1997) Molecular dynamics simulations of lipid bilayers. *Curr. Opin. Struct. Biol.* **7**:511 – 517.
- [38] Feller, S.E., Venable, R.M., Pastor, R.W. (1997) Computer simulation of a DPPC phospholipid bilayer: Structural changes as a function of molecular surface area. *Langmuir* **13**:6555 – 6561.
- [39] Scott, H.L. (2002) Modeling the lipid component of membranes. *Curr. Opin. Struct. Biol.* **12**:495 – 502.
- [40] Marrink, S.-J., Mark, A.E. (2001) Effect of undulations on surface tension in simulated bilayers. *J. Phys. Chem. B.* **105**:6122-6127.
- [41] Lindahl, E., Edholm, O. (2001) Molecular dynamics simulation of NMR relaxation rates and slow dynamics in lipid bilayers. *J. Chem. Phys.* **10**:4938 – 4950.
- [42] Saiz, L., Klein, M.L. (2002) Electrostatic interactions in a neutral model phospholipid bilayer by molecular dynamics simulations. *J. Chem. Phys.* **116**:3052 – 3057.
- [43] Wohlert, J., Edholm, O. (2004) The range and shielding of dipole-dipole interactions in phospholipid bilayers. *Biophys. J.* **87**:2433 – 2445.
- [44] Falck, E., Patra, M., Karttunen, M., Hyvonen, M.T., Vattulainen, I. (2004) Lessons of slicing membranes: Interplay of packing, free area, and lateral diffusion in phospholipid/cholesterol bilayers. *Biophys. J.* **87**:1076 – 1091.
- [45] Patra, M. (2005) Lateral pressure profiles in cholesterol-DPPC bilayers. *Eur. Biophys. J.* **35**:79 – 88.
- [46] Bemporad, D., Essex, J.W., Luttmann, C. (2004) Permeation of small molecules through a lipid bilayer: A computer simulation study. *J. Phys. Chem. B.* **108**:4875 – 4884.
-

-
- [47] Xiang, T.X., Anderson, B.D. (2006) Liposomal drug transport: A molecular perspective from molecular dynamics simulations in lipid bilayers. *Adv. Drug Deliver. Rev.* **58**:1357–1378.
- [48] Gullingsrud, J., Schulten, K. (2004) Lipid bilayer pressure proles and mechanosensitive channel gating. *Biophys. J.* **86**:3496–3509.
- [49] Carrillo-Tripp, M., Feller, S.E. (2005) Evidence for a mechanism by which omega-3 polyunsaturated lipids may affect membrane protein function. *Biochemistry* **44**:10164–10169.
- [50] Gullingsrud, J., Babakhani, A., McCammon, J.A. (2006) Computational investigation of pressure profiles in lipid bilayers with embedded proteins. *Mol. Simulat* **32**:831–838.
- [51] Murtola, T., Rog, T., Falck, E., Karttunen, M., Vattulainen, I. (2006) Transient ordered domains in single-component phospholipid bilayers. *Phys. Rev. Lett* **97**.
- [52] Anézo, C., Vries, A.H., Holtje, H.D., Tieleman, D.P., Marrink, S.-J. (2003) Methodological issues in lipid bilayer simulations. *J. Phys. Chem. B.* **107**:9424–9433.
- [53] Marrink, S.-J., Lindahl, E., Edholm, O., Mark, A.E. (2001) Simulation of the spontaneous aggregation of phospholipids into bilayers. *J. Am. Chem. Soc.* **123**:8638–8639.
- [54] Benz, R.W., Castro-Roman, F., Tobias, D.J., White, S.H. (2005) Experimental validation of molecular dynamics simulations of lipid bilayers: A new approach. *Biophys. J.* **88**:805–817.
- [55] de Vries, A.H., Chandrasekhar, I., Gunsteren, W.F., Hunenberger, P.H. (2005) Molecular dynamics simulations of phospholipid bilayers: Influence of artificial periodicity, system size, and simulation time. *J. Phys. Chem. B.* **109**:11643–11652.
- [56] Nielsen, S.O., Lopez, C.F., Srinivas, G., Klein, M.L. (2004) Coarse grain models and the computer simulation of soft materials. *J. Phys.: Condens. Mat.* **16**: R481–R512.
- [57] Shillcock, J.C., Lipowsky, R. (2006) The computational route from bilayer membranes to vesicle fusion. *J. Phys.: Condens. Mat.* **18**: S1191–S1219.
- [58] Muller, M., Katsov, K., Schick, M. (2006) Biological and synthetic membranes: What can be learned from a coarse-grained description? *Phys. Rep.* **434**:113–176.
- [59] Venturoli, M., Sperotto, M.M., Kranenburg, M., Smit, B. (2006) Mesoscopic models of biological membranes. *Phys. Rep.* **437**:1–54.
- [60] Smit, B., Hilbers, P.A.J., Esselink, K., Rupert, L.A.M., van Os, N.M., Schlijper, A.G. (1990) Computer simulations of a water/oil interface in the presence of micelles. *Nature* **348**:624–625.
-

- [61] Smit, B., Hilbers, P.A.J., Esselink, K., Rupert, L.A.M., van Os, N.M., Schlijper, A.G. (1991) Structure of a water/oil interface in the presence of micelles: a computer simulation study. *J. Phys. Chem.* **95**:6361 – 6368.
- [62] Groot, R.D., Warren, P.B. (1997) Dissipative particle dynamics: Bridging the gap between atomistic and mesoscopic simulation. *J. Chem. Phys.* **107**:4423 – 4435.
- [63] Frenkel, D., Smit, B. (2002) Understanding Molecular Simulation. Academic Press.
- [64] Venturoli, M., Smit, B. (1999) Simulating the self-assembly of model membranes. *Phys. Chem. Commun.* **10**:9/064721.
- [65] Groot, R.D. (2000) Mesoscopic simulation of polymer-surfactant aggregation. *Langmuir* **16**:7493 – 7502.
- [66] Groot, R.D., Rabone, K.L. (2001) Mesoscopic simulation of cell membrane damage, morphology change and rupture by nonionic surfactants. *Biophys. J.* **81**:725 – 736.
- [67] Kranenburg, M., Nicolas, J.P., Smit, B. (2004) Comparison of mesoscopic phospholipid-water models. *Phys. Chem. Chem. Phys.* **6**:4142 – 4151.
- [68] Kranenburg, M., Smit, B. (2005) Phase behavior of model lipid bilayers. *J. Phys. Chem. B.* **109**:6553 – 6563.
- [69] Groot, R.D. (2003) Electrostatic interactions in dissipative particle dynamics-simulation of polyelectrolytes and anionic surfactants. *J. Chem. Phys.* **118**:11265 – 11277.
- [70] Kranenburg, M., Venturoli, M., Smit, B. (2003) Phase behavior and induced interdigitation in bilayers studied with dissipative particle dynamics. *J. Phys. Chem. B.* **107**:11491 – 11501.
- [71] Kranenburg, M., Venturoli, M., Smit, B. (2003) Molecular simulations of mesoscopic bilayer phases. *Phys. Rev. E* **67**:060901.
- [72] Kranenburg, M., Laforge, C., Smit, B. (2004) Mesoscopic simulations of phase transitions in lipid bilayers. *Phys. Chem. Chem. Phys.* **6**:4531 – 4534.
- [73] Kranenburg, M., Smit, B. (2004) Simulating the effect of alcohol on the structure of a membrane. *FEBS Lett.* **568**:15 – 18.
- [74] Kranenburg, M., Vlaar, M., Smit, B. (2004) Simulating induced interdigitation in membranes. *Biophys. J.* **87**:1596 – 1605.
- [75] Venturoli, M., Smit, B., Sperotto, M.M. (2005) Simulation studies of protein-induced bilayer deformations, and lipid-induced protein tilting, on a mesoscopic model for lipid bilayers with embedded proteins. *Biophys. J.* **88**:1778 – 1798.
- [76] Shelley, J.C., Shelley, M.Y., Reeder, R.C., Bandyopadhyay, S., Klein, M.L. (2001) A coarse grain model for phospholipid simulations. *J. Phys. Chem. B.* **105**:4464 – 4470.
-

-
- [77] Shelley, J.C., Shelley, M.Y., Reeder, R.C., Bandyopadhyay, S., Moore, P.B., Klein, M.L. (2001) Simulations of phospholipids using a coarse grain model. *J. Phys. Chem. B.* **105**:9785 – 9792.
- [78] Lopez, C.F., Moore, P.B., Shelley, J.C., Shelley, M.Y., Klein, M.L. (2002) Computer simulation studies of biomembranes using a coarse grain model. *Comput. Phys. Commun.* **147**:1 – 6.
- [79] Pickholz, M., Saiz, L., Klein, M.L. (2005) Concentration effects of volatile anesthetics on the properties of model membranes: A coarse-grain approach. *Biophys. J.* **88**:1524 – 1534.
- [80] Srinivas, G., Klein, M.L. (2004) Computational approaches to nanobiotechnology: probing the interaction of synthetic molecules with phospholipid bilayers via a coarse grain model. *Nanotechnology* **15**:1289 – 1295.
- [81] Srinivas, G., Shelley, J.C., Nielsen, S.O., Discher, D.E., Klein, M.L. (2004) Simulation of diblock copolymer self-assembly, using a coarse-grain model. *J. Phys. Chem. B.* **108**:8153 – 8160.
- [82] Lopez, C.F., Nielsen, S.O., Moore, P.B., Klein, M.L. (2004) Understanding nature's design for a nanosyringe. *Proc. Natl Acad. Sci.* **101**:4431 – 4434.
- [83] Lopez, C.F., Nielsen, S.O., Ensing, B., Moore, P.B., Klein, M.L. (2005) Structure and dynamics of model pore insertion into a membrane. *Biophys. J.* **88**:3083 – 3094.
- [84] Nielsen, S.O., Ensing, B., Ortiz, V., Moore, P.B., Klein, M.L. (2005) Lipid bilayer perturbations around a transmembrane nanotube: A coarse grain molecular dynamics study. *Biophys. J.* **88**:3822 – 3828.
- [85] Nielsen, S.O., Lopez, C.F., Ivanov, I., Moore, P.B., Shelley, J.C., Klein, M.L. (2004) Transmembrane peptide-induced lipid sorting and mechanism of L_{α} -to-inverted phase transition using coarse-grain molecular dynamics. *Biophys. J.* **87**:2107 – 2115.
- [86] Lopez, C.F., Nielsen, S.O., Srinivas, G., Degrado, W.F., Klein, M.L. (2006) Probing membrane insertion activity of antimicrobial polymers via coarse-grain molecular dynamics. *J. Chem. Theory Comput.* **2**:649 – 655.
- [87] Marrink, S.-J., Vries, A.H., Mark, A.E. (2004) Coarse grained model for semiquantitative lipid simulations. *J. Phys. Chem. B.* **108**:750 – 760.
- [88] Marrink, S.-J., Risselada, J., Mark, A.E. (2005) Simulation of gel phase formation and melting in lipid bilayers using a coarse grained model. *Chem. Phys. Lipids* **135**:223 – 244.
- [89] Marrink, S.-J., Mark, A.E. (2003) Molecular dynamics simulation of the formation, structure, and dynamics of small phospholipid vesicles. *J. Am. Chem. Soc.* **125**:15233 – 15242.
-

- [90] Marrink, S.-J., Mark, A.E. (2003) The mechanism of vesicle fusion as revealed by molecular dynamics simulations. *J. Am. Chem. Soc.* **125**:11144–11145.
- [91] de Joannis, J., Jiang, F.Y., Kindt, J.T. (2006) Coarse-grained model simulations of mixed-lipid systems: Composition and line tension of a stabilized bilayer edge. *Langmuir* **22**:998–1005.
- [92] Marrink, S.-J., Mark, A.E. (2004) Molecular view of hexagonal phase formation in phospholipids membranes. *Biophys. J.* **87**:3894-3900.
- [93] Faller, R., Marrink, S.-J. (2004) Simulation of domain formation in DLPC-DSPC mixed bilayers. *Langmuir* **20**:7686–7693.
- [94] Shi, Q., Voth, G.A. (2005) Multi-scale modeling of phase separation in mixed lipid bilayers. *Biophys. J.* **89**:2385–2394.
- [95] Dickey, A.N., Faller, R. (2005) Investigating interactions of biomembranes and alcohols: A multiscale approach. *J. Polym. Sci.* **43**:1025–1032.
- [96] Lee, H., Larson, R.G. (2006) Molecular dynamics simulations of PAMAM dendrimer-induced pore formation in DPPC bilayers with a coarse-grained model. *J. Phys. Chem. B.* **110**:18204–18211.
- [97] Bond, P.J., Sansom, M.S.P. (2006) Insertion and assembly of membrane proteins via simulation. *J. Am. Chem. Soc.* **128**:2697–2704.
- [98] Shih, A., Arkhipov, A., Freddolino, P., Schulten, K. (2006) Coarse grained protein-lipid model with application to lipoprotein particles. *J. Phys. Chem. B.* **110**:3674–3684.
- [99] Adhangale, P.S., Gaver, D.P. (2006) Equation of state for a coarse-grained DPPC monolayer at the air/water interface. *Mol. Phys.* **104**:3011–3019.
- [100] Izvekov, S., Voth, G.A. (2005) A multiscale coarse-graining method for biomolecular systems. *J. Phys. Chem. B.* **109**:2469–2473.
- [101] Izvekov, S., Voth, G.A. (2006) Multiscale coarse-graining of mixed phospholipid/cholesterol bilayers. *J. Chem. Theory Comput.* **2**:637–648.
- [102] Shi, Q., Izvekov, S., Voth, G.A. (2006) Mixed atomistic and coarse-grained molecular dynamics: Simulation of a membrane-bound ion channel. *J. Phys. Chem. B.* **110**:15045–15048.
- [103] Goetz, R., Lipowsky, R. (1998) Computer simulations of bilayer membranes: Self-assembly and interfacial tension. *J. Chem. Phys.* **108**:7397–7409.
- [104] Goetz, R., Gompper, G., Lipowsky, R. (1999) Mobility and elasticity of self-assembled membranes. *Phys. Rev. Lett.* **82**:221–224.
- [105] Imparato, A., Shillcock, J.C., Lipowsky, R. (2003) Lateral and transverse diffusion in two-component bilayer membranes. *Eur. Phys. J.E.* **11**:21–28.
-

-
- [106] Imparato, A., Shillcock, J.C., Lipowsky, R. (2005) Shape fluctuations and elastic properties of two-component bilayer membranes. *Europhys. Lett.* **69**:650-656.
- [107] den Otter, W.K., Briels, W.J. (2003) The bending rigidity of an amphiphilic bilayer from equilibrium and nonequilibrium molecular dynamics. *J. Chem. Phys.* **118**:4712 – 4720.
- [108] Helfrich, W. (1973) Elastic properties of lipid bilayers -theory and possible experiments. *Z. Naturforsch.* **28c**:693 – 703.
- [109] Boek, E.S., den Otter, W.K., Briels, W.J., Iakovlev, D. (2004) Molecular-dynamics simulation of amphiphilic bilayer membranes and wormlike micelles: a multi-scale modelling approach to the design of viscoelastic surfactant solutions. *Phil. Trans. R. Soc. Lond. A.* **362**:1625 – 1638.
- [110] Loison, C., Mareschal, M., Kremer, K., Schmid, F. (2003) Thermal fluctuations in a lamellar phase of a binary amphiphile-solvent mixture: A molecular-dynamics study. *J. Chem. Phys.* **119**:13138 – 13148.
- [111] Loison, C., Mareschal, M., Schmid, F. (2004) Pores in bilayer membranes of amphiphilic molecules: Coarse-grained molecular dynamics simulations compared with simple mesoscopic models. *J. Chem. Phys.* **121**:1890-1900.
- [112] Loison, C., Mareschal, M., Schmid, F. (2005) Fluctuations and defects in lamellar stacks of amphiphilic bilayers. *Comput. Phys. Commun.* **169**:99 – 103.
- [113] Stevens, M.J. (2004) Coarse-grained simulations of lipid bilayers. *J. Chem. Phys.* **121**:11942 – 11948.
- [114] Stevens, M.J., Hoh, J.H., Woolf, T.B. (2003) Insights into the molecular mechanism of membrane fusion from simulation: Evidence for the association of splayed tails. *Phys. Rev. Lett.* **91**:188102.
- [115] Lenz, O., Schmid, F. (2005) A simple computer model for liquid lipid bilayers. *J. Mol. Liquid.* **117**:147 – 152.
- [116] Brannigan, G., Lin, L.C.L., Brown, F.L.H. (2006) Implicit solvent simulation models for biomembranes. *Eur. Biophys. J.* **35**:104 – 124.
- [117] Whitehead, L., Edge, C.M., Essex, J.W. (2001) Molecular dynamics simulation of the hydrocarbon region of a biomembrane using a reduced representation model. *J. Comput. Chem.* **22**:1622 – 1633.
- [118] Gay, J.G., Berne, B.J. (1981) Modification of the overlap potential to mimic a linear site-site potential. *J. Chem. Phys.* **74**:3316 – 3319.
- [119] Noguchi, H., Takasu, M. (2001) Self-assembly of amphiphiles into vesicles: A Brownian dynamics simulation. *Phys. Rev. E.* **64**:041913.
- [120] Noguchi, H., Takasu, M. (2001) Fusion pathways of vesicles: A Brownian dynamics simulation. *J. Chem. Phys.* **115**:9547 – 9551.
-

- [121] Noguchi, H., Takasu, M.(2002) Adhesion of nanoparticles to vesicles: A Brownian dynamics simulation. *Biophys. J.* **83**:299 – 308.
 - [122] Noguchi, H., Takasu, M.(2002) Structural changes of pulled vesicles: A Brownian dynamics simulation. *Phys. Rev. E.* **65**:051907.
 - [123] Lyubartsev, A.P. (2005) Multiscale modeling of lipids and lipid bilayers. *Eur. Biophys. J.* **35**:53 – 61.
 - [124] Lyubartsev, A.P., Laaksonen, A. (2004) On the reduction of molecular degrees of freedom in computer simulations. *Lecture Notes Phys.* **640**:214 – 244.
 - [125] Brannigan, G., Brown, F.L.H. (2004) Solvent-free simulations of fluid membrane bilayers. *J. Chem. Phys.* **120**:1059 – 1071.
 - [126] Brannigan, G., Tamboli, A.C., Brown, F.L.H. (2004) The role of molecular shape in bilayer elasticity and phase behavior. *J. Chem. Phys.* **121**:3259 – 3271.
 - [127] Brannigan, G., Philips, P.F., Brown, F.L.H. (2005) Flexible lipid bilayers in implicit solvent. *Phys.Rev.E.* **72**:011915.
 - [128] Brannigan, G., Brown, F.L.H. (2005) Composition dependence of bilayer elasticity. *J. Chem.Phys.* **122**:074905.
-

