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The STRENDA Commission (<u>St</u>andards for <u>Reporting Enzymology Da</u>ta) compiled the following Guidelines, as a service to the community, to define the minimum amount of information that should accompany any published enzyme activity data.

The current STRENDA Guidelines (List Level 1B) was reviewed on the STRENDA meeting in November 2021 in terms of consistency of form and content, as well as of the order and plausibility of the list entries. In addition, it now includes recommendations for reporting on the accuracy and deposition of data as well as those parameters which should be reported when equilibrium measurements have been performed.

List Level 1B

defines those data that are required to allow a quality check on the data and to ensure their value to others. In principle, this is the minimum information to describe enzyme activity data.

| Information required | Comments |
|--------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Required data for all enzyme functional data | |
| Reproducibility, number of independent experiments | Indicate how many times the measurement was reproduced and what changed between replicates; just repeat reactions, different enzyme preparations, different ways, alternative staff, different laboratories |
| Precision of measurement | e.g., standard error of the mean, standard deviation, confidence limits, quartiles. |
| | Comments on possible systematic errors are also useful. |
| Specification whether relative to subunit or oligomeric form | |





| Information required | Comments |
|----------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Preferably deposit measured/raw data (e.g., time course of product concentrations) to enable re-analysis | Make data findable by citing DOI or URL, accessible by making it open, interoperable by structuring and describing the data by using formats such as EnzymeML |
| Data necessary for reporting kinetic parameters | (choose which ones are available from your experiments) |
| Kinetic equation (which will then define parameters) | Name or state the model or equation used and the variable in it, e.g., Michaelis-Menten, varying concentration of ATP, fixed glucose or $v = V_{max} / (1 + K_A/[2-aminopropane] + K_B/[2-butanone])$ |
| <i>k</i> _{cat} | V_{\max} in terms of mol reaction per mol enzyme per time, so units often reported as s ⁻¹ or min ⁻¹ |
| V _{max} | Should be as a specific activity, with units like mol min ⁻¹ (g enzyme) ⁻¹ , or (mol product) min ⁻¹ (g protein in the preparation) ⁻¹ , <i>see</i> List Level 1A |
| $k_{ m cat}/K_{ m m}$ | $k_{\rm cat}/K_{ m m}$ given as per concentration per time |
| | e.g., mM ⁻¹ s ⁻¹ |
| $K_{ m m}$ | units or concentration necessary, e.g., mM, |
| | define how $K_{\rm m}$ was defined operationally (e.g. as $S_{0.5}$) |
| $\mathcal{S}_{0.5}$ | concentration, e.g., mM |
| Hill coefficient, saturation ratio (RS) or other coefficients of cooperativity | with equation defining the parameter as noted above |
| How was the given parameter obtained? | e.g., non-linear curve fitting using least squares, non- parametric method such as direct linear plot, linear regression to transformed form of rate equation. |
| | Note: if commercial computer programs are used, determine which were used |
| K _{M2} | Michaelis constants for all co-substrates, inclusive the coenzymes |





| Information required | Comments |
|--------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| K _P | $K_{\rm m}$ for reverse operation or product inhibition constants (with equation above showing definition). This is for all products inclusive of cofactors. If information is absent, indicate at what product concentrations there was no effect on enzyme rate. |
| Choice of model used to determine the parameters | Report any measures of quality of fit, and for any alternative models considered. |
| High-substrate inhibition, if observed, with <i>K</i> _i value | with defining equation above |
| Data required for reporting inhibition data | |
| Time-dependence and reversibility | with method described |
| Inhibition | <i>K</i> _i units necessary |
| types: | |
| reversible | e.g., competitive, uncompetitive, etc., with units and how values were determined |
| tight-binding | association/dissociation rates; estimates OK if small |
| irreversible | e.g. non-specific, mechanism-based, "suicide substrate". |
| | There are too many alternative parameters to list here. The reference to a quite comprehensive source is recommended: Enzymes: Irreversible Inhibition. McDonald, A.G. & Tipton, K.F. In: Nature Encyclopedia of Life Sciences London (2020). doi:10.1002/9780470015902.a0000601.pub3 |
| | Note: <i>IC</i> ₅₀ values |
| | These have been used for both reversible or irreversible inhibition. However, the use is not recommended because these values are without a consistent meaning. The relationship of these values to inhibition constants is analysed in details, e.g., by Cortes, A. <i>et al.</i> (2001) <i>Biochem. J.</i> 357 :263-268. doi:10.1042/bj3570263 |
| Data required for reporting activation data | similar to the requirements for inhibition data |





| Information required | Comments |
|----------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Data required for reporting | See further details in IUBMB document: |
| Data required for reporting equilibrium measurement | Alberty R.A. <i>et al.</i> (2011) <i>Biophys. Chem.</i> 155 :89-103. <u>doi:10.1016/j.bpc.2011.03.007</u> |
| Measured equilibrium concentrations | preferred that these are tabulated |
| K _{eq} ' or K' (i.e. the pH dependent equilibrium constant) | with reference to full reaction equation presented and direction identified. |
| | with units where not symmetrical, e.g. M, mM ⁻¹ , mol kg ⁻¹ (molality). Explain any reactants not treated by way of dissolved concentrations, e.g., water, gases as partial pressures, activities for reactants not behaving as infinitely dilute. |
| | Estimates of equilibrium constants may sometimes also be obtained by fitting to kinetic (progressive) data. If so, follow the recommendations as for other kinetic parameters, including stating the equation fitted. |
| Convention used | by default assume biochemical convention using total concentrations of ionising or complexing compounds. But state clearly if using defined chemical species. |

About the STRENDA Commission:

The STRENDA Commission is formed by an international panel of highly-regarded scientists who bring in diverse expertises such as biochemistry, enzyme nomenclature, bioinformatics, systems biology, modelling, mechanistic enzymology and theoretical biology.

The Commission was founded in 2003 and is supported by the Beilstein-Institut since then.

Members of the Commission are: **B.M. Bakker** (University Medical Center Groningen, The Netherlands), **A. Cornish-Bowden** (CNRS-BIP, Marseilles, France), **P. Fitzpatrick** (University of Texas Health Science Center at San Antonio, San Antonio, TX, USA), **P. Halling** (University of Strathclyde, Glasgow, UK), **T.S. Leyh** (The Albert Einstein College of Medicine, Bronx, NY, USA), **A.G. McDonald** (Trinity College Dublin, Ireland), **M. Neumann-Schaal** (Leibniz Institute DSMZ, Braunschweig, Germany), **C. O'Donovan** (EBI, Cambridge, UK), **J. Pleiss** (University of Stuttgart, Germany), **F.M.Raushel** (Texas A&M University, College Station, TX, USA), **J.M. Rohwer** (University of Stellenbosch, South Africa), **S. Schnell** (University of Notre Dame, IN, USA), **N. Swainston** (The University of Liverpool, UK), **M.-D. Tsai** (Academia Sinica, Taipeh, Taiwan), **K. Tipton** (Trinity College, Dublin, Ireland), **H.V. Westerhoff** (Universities of Amsterdam, The Netherlands),





U. Wittig (Heidelberg Institute of Theoretical Studies, Germany) **R. Wohlgemuth** (Lodz University of Technology, Poland) and **C. Kettner** (co-ordination, Beilstein-Institut, Frankfurt, Germany).

More information: <u>www.beilstein-strenda.org</u>